Spatial Distribution and Binomial Sampling of *Aphis gossypii* Glover (Homoptera: Aphididae) Infesting Protected Cucumber and Melon in Northern Italy. (*)

**INTRODUCTION**

*Aphis gossypii* Glover, the main pest of protected cucurbits in Italy (Celli et al., 1991) and northern Europe (van Lenteren & Woets, 1988), is also becoming an increasing threat to other greenhouse crops like sweet pepper. The development of strains resistant to pirimicarb is a serious drawback to the use of beneficial arthropods, and many countries are attempting to develop biological control strategies (van Steenis, 1992).

Studying the spatial distribution pattern of a pest is the first step to designing a sampling method. Common approaches in entomology are determinations of mean-variance relationships (Taylor, 1961; 1984), frequency distribution (Anscombe, 1949; Bliss & Owen, 1958), mean crowding (Lloyd, 1967; Iwao, 1968) or, more recently, a new survey based on an improvement of Iwao’s model (Xu et al., 1993). Other techniques, like geostatistics, determine the degree of association (correlation) among samples based on the direction and distance between them (Schotzko & Knudsen, 1992). The latter is a kind of analysis that quantitatively evaluates variations or changes in spatial orientation within a defined area or volume. The present study reports two-year of data for *A. gossypii* in cucumber and melon crops of northern Italy’s Po Valley in an attempt to determine viable sampling strategies for scouting and research of this aphid.

**MATERIALS AND METHODS**

1992. The samples of *A. gossypii* were taken in cucumber tunnels (250 m² each), two at Granarolo (Bologna Province) and four at Cesena (Forlì Province), from early May to July-August, with the end date depending on site and infestation severity. The number of aphids per leaf and their development stage were recorded weekly on 200 randomly selected leaves of 100 plants.

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1993. These samples involved two 300 m² tunnels of the cucumber cv. ‘Darina’ at Granarolo and four 250 m² tunnels of melon at San Pietro in Casale (Bologna Province). The cucumber data were recorded as in 1992, although leaf number that year ranged from 150 to 400; the melon count included 10-12 leaves per plant, for a total of 400 leaves.

Statistical analysis. The mean number of A. gossypii per leaf (m) and the variance (s²) were calculated for total leaves sampled per tunnel and per sampling date. Taylor’s power law (s² = amᵇ) (Taylor, 1961; 1984), which describes the correlation of means to sampling variances, was employed to study the spatial distribution of A. gossypii, the estimates of parameters (a and b) being calculated by regression of log (s²) over log (m) to yield:

\[
\log (s^2) = \log a + b \log (m),
\]

(1)

where the intercept (a) is a parameter essentially dependent on sampling method and the angular coefficient (b) is defined as the index of aggregation; as such the latter is a constant per species and varies continuously from a regular distribution for b → 0, to random for b = 1, to clumped for b > 1 (Taylor, 1961). To calculate the optimum sample size (OSS) of A. gossypii for direct count, the sampling variance from Taylor’s power law was introduced in Karandinos’s formula (1976),

\[
n = \left(\frac{Z_{a/2}}{D}\right)^2 \frac{s^2}{m^2},
\]

(2)

where D is the required level of accuracy expressed as a decimal (0.2-0.3-0.5) or permitted percentage of error, and Zα/2 is the standard normal deviate; for n > 30 and with α = 0.05, Zα/2 = 1.96 (Karandinos, 1976).

The model of Gerrard & Chiang (1970) was used to predict mean aphid density (m) by the proportion of empty sample units (p₀):

\[
\ln(m) = \alpha + \beta \ln [- \ln(p_0)],
\]

(3)

where \(\alpha\) and \(\beta\) are constants. This equation yields

\[
m = e^{\alpha} [- \ln (p_0)]^\beta
\]

(4)

(Binns & Bostonian, 1990). The variance of a predicted mean is the sum of a prediction variance [varp(m)] and a sampling variance [vars(m)] (Nyrop & Binns, 1991). The prediction variance is

\[
\text{varp(m)} = m^2 \left[ \frac{\text{mse}}{N} + \ln [- \ln (p_0)] - \text{avglnln}(p_0) \right]^2 s^2 + \text{mse},
\]

(5)

where \(\text{mse}\) is the mean square error from the regression, \(N\) is the number of data points in the regression, \(\text{avglnln}(p_0)\) is the average of the independent variable in the regression and \(s^2\) is the variance of the regression slope. The sampling variance is

\[
\text{vars(m)} = p_0 (1 - p_0) \frac{s^2 m}{n},
\]

(6)

where \(p\) is the slope of the relationship between \(m\) and \(p_0\) as per Eq. (3), and \(n\) is the sample size to estimate \(m\) from \(p_0\).

Analysis of covariance (ANCOVA) was used to compare the parameters of Taylor’s power law, i.e. to determine if the regression for year, crop and location were coincidental or not. The comparison between year and location for cucumber was made first and then that between cucumber and melon.
RESULTS AND DISCUSSION

The fit of Taylor's model to the data per crop and year is shown in Tab. 1: the high coefficients of determination ($r^2$ ranged between 0.97 and 0.98) for the relative regression equations indicate the good fit of the model. Covariance analysis found no significant differences between the coefficients of Taylor's power law for cucumber at the two sites and years (Tab. 2); the regressions for cucumber were coincidental and a common regression was used to determine the relationship between mean and variance in this crop (Tabs. 1, 2; Fig. 1). The regression slopes showed some variability for cucumber between 1992 and 1993, and the parallelism test showed a value of $P = 0.09$, which is near the level of significance and

Tab. 1 - Coefficients from Taylor's power law calculated for A. gossypii per crop and year.

<table>
<thead>
<tr>
<th>Crop</th>
<th>Location</th>
<th>Years</th>
<th>n</th>
<th>log a ($\pm$s.e.)</th>
<th>b ($\pm$s.e.)</th>
<th>$r^2$</th>
<th>P</th>
</tr>
</thead>
<tbody>
<tr>
<td>Cucumber</td>
<td>Granarolo (BO)</td>
<td>1992</td>
<td>17</td>
<td>1.63 (0.06)</td>
<td>1.82 (0.06)</td>
<td>0.98</td>
<td>&lt; 0.001</td>
</tr>
<tr>
<td>Cucumber</td>
<td>Cesena (FO)</td>
<td>1992</td>
<td>17</td>
<td>1.75 (0.06)</td>
<td>1.78 (0.07)</td>
<td>0.97</td>
<td>&lt; 0.001</td>
</tr>
<tr>
<td>Cucumber</td>
<td>Granarolo (BO)</td>
<td>1993</td>
<td>23</td>
<td>1.72 (0.06)</td>
<td>1.63 (0.04)</td>
<td>0.98</td>
<td>&lt; 0.001</td>
</tr>
<tr>
<td>Cucumber</td>
<td>Cesena (FO)</td>
<td>1993</td>
<td>23</td>
<td>1.70 (0.04)</td>
<td>1.71 (0.03)</td>
<td>0.97</td>
<td>&lt; 0.001</td>
</tr>
<tr>
<td>Melon</td>
<td>S.Pietro in Casale (BO)</td>
<td>1993</td>
<td>22</td>
<td>1.94 (0.06)</td>
<td>1.66 (0.04)</td>
<td>0.98</td>
<td>&lt; 0.001</td>
</tr>
</tbody>
</table>

Tab. 2 - Comparison of coefficients of Taylor's power law by analysis of covariance for A. gossypii for cucumber per years and locations.

<table>
<thead>
<tr>
<th>Location and year</th>
<th>n</th>
<th>Intercept (log a)</th>
<th>F</th>
<th>P</th>
<th>Slope (b)</th>
<th>F</th>
<th>P</th>
</tr>
</thead>
<tbody>
<tr>
<td>BO - 1992</td>
<td>17</td>
<td>1.63</td>
<td>1.82</td>
<td>0.228</td>
<td>1.78</td>
<td>2.54</td>
<td>0.088</td>
</tr>
<tr>
<td>FO - 1992</td>
<td>17</td>
<td>1.75</td>
<td>1.52</td>
<td>0.028</td>
<td>1.78</td>
<td>1.63</td>
<td>0.63</td>
</tr>
<tr>
<td>BO - 1993</td>
<td>23</td>
<td>1.72</td>
<td>1.63</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

Tab. 3 - Comparison of coefficients of Taylor's power law by analysis of covariance for A. gossypii in cucumber (data pooled for all sites and years) and melon.

<table>
<thead>
<tr>
<th>Location and year</th>
<th>n</th>
<th>Intercept (log a)</th>
<th>F</th>
<th>P</th>
<th>Slope (b)</th>
<th>F</th>
<th>P</th>
</tr>
</thead>
<tbody>
<tr>
<td>Cucumber (common)</td>
<td>57</td>
<td>1.70</td>
<td>4.71</td>
<td>0.003</td>
<td>0.73</td>
<td>0.39</td>
<td></td>
</tr>
<tr>
<td>Melon</td>
<td>22</td>
<td>1.94</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

Tab. 4 - Parameters from the regression of ln (m) on ln (- ln(p)) for cucumber and melon to estimate aphid density by incidence of infestation (for the symbols, see the test).

<table>
<thead>
<tr>
<th>Crop</th>
<th>$\alpha$ ($\pm$s.e.)</th>
<th>$\beta$ ($\pm$s.e.)</th>
<th>$r^2$</th>
<th>P</th>
<th>$mse$</th>
<th>$sB^2$</th>
<th>N</th>
</tr>
</thead>
<tbody>
<tr>
<td>Cucumber</td>
<td>4.11 (0.22)</td>
<td>1.45 (0.08)</td>
<td>0.86</td>
<td>&lt; 0.001</td>
<td>- 2.218</td>
<td>0.994</td>
<td>0.006</td>
</tr>
<tr>
<td>Melon</td>
<td>4.86 (0.39)</td>
<td>1.49 (0.11)</td>
<td>0.89</td>
<td>&lt; 0.001</td>
<td>- 2.810</td>
<td>1.14</td>
<td>0.012</td>
</tr>
</tbody>
</table>
Cucumber (common)

Fig. I - Taylor’s power law for *A. gossypii* on cucumber (data pooled).

Melon

Fig. II - Taylor’s power law applied to *A. gossypii* infestation of melon in Bologna Province (1993).
Fig. III - Numerical sample size curves for *A. gossypii* on cucumber at three confidence levels (*D* = 0.2, 0.3, 0.5, *α* = 0.05).

Fig. IV - Numerical sample size curves for *A. gossypii* on melon at three confidence levels (*D* = 0.2, 0.3, 0.5, *α* = 0.05).
Fig. V - Population mean density \( Y \) of \textit{A. gossypii} on cucumber \( Y_1 \) and melon \( Y_2 \) as estimated by equations \( Y_1 = e^{3.6 - \ln p_0^{1.45}} \), \( Y_2 = e^{3.3 - \ln p_0^{1.39}} \).

Fig. VI - Variances for \textit{A. gossypii} density estimates from a sample of 400 leaves. For the variances, see Eqs. (3) and (6).
may be attributable to a certain variability in the type of infestation exhibited by
*A. gossypii*. Ants that are symbiotic with this aphid play an important role in its
type of infestation, and hence in the variation in the data. Two species of ants, *Formica
cunicularia* Latr. and *F. cinerea* Mayr were present near the aphid colonies
throughout the sampling periods.

The comparison of slope coefficients of Taylor’s power law between cucumber
(pool data) and melon showed no statistical difference (Tabs. 1, 2, 3; Figs. 1, II).
This lack of a difference is in accordance with Taylor’s theory which postulates
that b is a constant for a species. The value of b for both plant species exhibits a
high degree of aggregation for *A. gossypii*. However, the two intercepts showed a
statistical difference (Tab. 3). This difference is logical because it incorporates the
effects of sample unit and host crop (Taylor, 1984). Thus, although from a biological
viewpoint a single index of aggregation could be used as typical for the species
so as to calculate the OSS in cucumber and melon, it is definitely more cor-
rect to use the parameters a and b of the separate equations (Tab. 1).

Guldemond (1993), using Taylor’s law, calculated an angular coefficient of 1.43
for *A. gossypii* on cut chrysanthemums in greenhouse, which is a lower value for
b than reported in the present study. It would be interesting to determine whether
this difference results from different strains of *A. gossypii* in Northern Europe and
the Mediterranean region. Though b can frequently be considered a species-spe-
cific constant (Taylor, 1961), it can vary markedly for some aphid species (Elliott &
Kieckhefer, 1987) and can be influenced by genetic differences, influencing
life-history traits (Taylor, 1984).

The curves plotted for the numerical sample size as calculated by Eq. (6) are
shown in Figs. III and IV. When a higher accuracy is required, more samples have
to be taken. Yet for a level of accuracy of D = 0.2, the number of samples of lea-
ves proved to be too high and not applicable in actual practice. Feng & Nowierski
(1992) advise that for scouting purposes both extension agents and growers use an
accuracy of D = 0.5, whereas D should be at least 0.3 for research purposes.
In our case, for example, estimating a population at 50, 100, 150 aphids per leaf
requires, respectively, 723, 596 and 532 leaves for cucumber and 989, 783 and
682 for melon at D = 0.3, whereas the number of leaves required for the same
population levels in a practical situation, i.e. for an accuracy of D = 0.5, are 260,
214 and 191 for cucumber and 356, 281 and 245 for melon.

Fig. V shows the regressions calculated by Eq. (6), which estimates aphid density
via the incidence of uninfested leaves, and Table 4 lists the matching parameters. To
correct for the bias introduced by transforming estimates of ln(m) back to original
values, half of the residual mean square from the regression was added to the inter-
cept (Hepworth & MacFarlane, 1992). Thus, in going from the logarithms to the or-
ginal values and keeping this fact in mind, the aphid population can be estimated by
the frequency of uninfested leaves as per Eq. (6) (see Fig. V), the variance of m being
calculated with Eqs. (6) and (6) (Fig. VI) (Nyrop & Binns, 1991). However, because of the
low intervention thresholds for cucurbit crops, the use of binomial sampling for
*A. gossypii* on cucurbits could be somewhat problematic and dangerous, compared to
other crop-pest systems (Ekholm, 1987; Binns & Bostonian, 1991; Nyrop & Binns,
The biological control techniques now under study, such as releases of the aphid parasitoid Aphidius colemani Viereck or of the predator Aphidolates aphidi-mize Rond., require that the introductions of these beneficials either be timed to the onset of aphid attack (van Stenis, 1992) or even precede natural infestation through the use of banker plants (Bennison, 1992) or the AHMAF method (Stary, 1993). There is also a certain variability in the data, especially if the density of the aphids is estimated on the basis of low infestation rates (low number of infested leaves). The estimate of the aphid’s population density is useful, however, in certain cases, e.g. to assess the results of IPM or biological control trials and, in general, whenever an estimate of infestation level is needed and one is short of both time and labour.

KEY WORDS: Aphis gossypii, cucumber, melon, spatial distribution, binomial sampling

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SUMMARY

Taylor’s power law was employed to study the spatial distribution of Aphis gossypii in tunnel-grown cucumber and melon in northern Italy’s Emilia-Romagna in order to improve sampling procedures. The aggregation index (h) in cucumber showed no significant differences between years (1992 and 1993), sites (Bologna and Cesena) or crop (h = 1.66 for melon and 1.71 for cucumber). Binomial sampling, which correlates infestation severity to infested leaf frequency, is also reported and discussed, along with problems inherent in sampling and in control methods for A. gossypii that are linked to the peculiar aggregate type of distribution of this aphid in the studied crops and the rapid development of infestations in field.

Distribuzione spaziale e campionamento binomiale di Aphis gossypii Glover (Homoptera: Aphididae) infestante cetriolo e melone in coltura protetta in Emilia-Romagna.

RIASSUNTO

È stata utilizzata la legge di Taylor nello studio della distribuzione spaziale di Aphis gossypii Glover in tunnel di cetriolo e melone in Emilia-Romagna. L’indice di aggregazione (h) su cetriolo non mostrò differenze significative riguardo sia alle due annate di studio (1992-1993) che alle due diverse località (Bologna-Cesena), non furono inoltre evidenziate differenze significative nel tipo di aggregazione fra cetriolo (h = 1.71) e melone (h = 1.66). Viene presentato un campionamento di tipo binomiale, che permette di stimare la densità delle popolazioni dell’afide, dalla frequenza delle foglie infestate. Sono messe in rilievo le problematiche del campionamento e della lotta contro tale fito-
go, legate alla sua distribuzione spaziale particolarmente aggregata, al rapido sviluppo in coltura protetta, ed alla bassa soglia economica.

REFERENCES CITED


